# Prevalence of Cryptosporidiosis and Molecular Characterization of *Cryptosporidium* spp. in Calves in Erzurum<sup>[1]</sup>

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### Summary

This study was conducted to determine the prevalence of cryptosporidiosis and to identify *Cryptosporidium* species found in preweaned calves, in Erzurum, Turkey. Fecal samples were collected from 307 calves up to one month old from 5 dairy farms. Genomic DNA was obtained by DNA extraction (QIAamp DNA Stool kit). The prevalence of cryptosporidiosis was determined based on identification through a nested PCR protocol to amplify fragments of the *Cryptosporidium* SSU rRNA gene. 3.9% of calves were positive for *Cryptosporidium*. Calves that were subjected to traditional herd management, were female, aged 2 weeks, and had watery feces were affected by the disease at a greater incidence than those were subjected to planned herd surveillance program, were males, were older than 3 weeks, and had firm feces. DNA sequence analysis of the SSU rRNA gene on all of the PCR positive samples ascertained that *C. parvum* was the only species present. Further studies should be performed comprehensive fecal analysis for other causative agents for association *Cryptosporidium* species in calf diarrhea and mortality resulting in economic loss in the region.

Keywords: Calf, Cryptosporidium, Nested-PCR, SSU rRNA, Erzurum

# Erzurum Yöresinde Buzağılarda Cryptosporidiosisin Prevalansı ve *Cryptosporidium* Türlerinin Moleküler Karekterizasyonu

## Özet

Bu çalışma, Erzurum yöresindeki sütten kesim öncesi dönemdeki buzağılarda cryptosporidiosisin prevalansının ve *Cryptosporidium* türlerinin moleküler karekterizasyonunun ortaya konması amacıyla yapılmıştır. Bu amaçla beş süt işletmesinden, bir aydan küçük 307 buzağının dışkı örnekleri toplanmış ve DNA ekstraksiyonu (QIAamp DNA Stool kit) yapılarak genomik DNA elde edilmiştir. Nested PCR protokolü ile *Cryptosporidium* SSU rRNA gen bölgesinin kısmi amplifikasyonu yapılmış ve cryptosporidiosis prevalansı %3.9 olarak belirlenmiştir. Geleneksel yöntemlerle yetiştirilen, dişi, 2 haftalık yaşta ve sulu dışkıya sahip buzağıların modern işletmelerde yetiştirilen, erkek, 3 haftadan büyük ve katı kıvamlı dışkıya sahip olanlara göre hastalıktan daha çok etkilendiği saptanmıştır. PCR pozitif örneklerin SSU rRNA gen bölgesi hedef alınarak yapılan DNA dizi analizleri sonuçlarına göre *C. parvum*'un hayvanlarda bulunan tek tür olduğu anlaşılmıştır. Sonuç olarak, yörede ekonomik kayıplara neden olan buzağı ishalleri ve ölümlerinde *Cryptosporidium* türleri ile diğer hastalık etkenlerinin etkileşimlerinin ortaya konması amacıyla daha kapsamlı çalışmaların yapılmasının gerekliliği sonucuna varılmıştır.

Anahtar sözcükler: Buzağı, Cryptosporidium, Nested-PCR, SSU rRNA, Erzurum

## INTRODUCTION

Cryptosporidiosis is a zoonotic protozoan disease that has a very broad and versatile geographic distribution including the Antarctic region <sup>[1]</sup>. *Cryptosporidium* is the

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causative agent and infects mainly the intestinal tract and rarely the respiratory system of diverse species including human, ruminant, feline, canine, rodent, avian, reptile and fish. Transmission usually occurs through the direct fecal-oral route or through ingestion of water or food contaminated with oocysts <sup>[2,3]</sup>.

Bovines are the most common species of mammals, infected with *Cryptosporidium* and considered the major reservoir of *Cryptosporidium* for human infections <sup>[3]</sup>. Cryptosporidiosis in cattle is mainly caused by *C. parvum*, *C. andersoni, C. bovis* and *C. ryanae* <sup>[2,3]</sup>. At least 10 other *Cryptosporidium* species or genotypes such as *C. felis, C. meleagridis* and *C. suis* can also play role in etiology <sup>[4,5]</sup>.

Bovine cryptosporidiosis is considered one of the most common causes of neonatal diarrhea in cattle, leading to economic losses. The severity of infection ranges from mild to severe depending on *Cryptosporidium* species as well as age, previous exposure and immune status of the host. Asymptomatic infection is common in yearling heifers and mature cows <sup>[6,7]</sup>.

*Cryptosporidium parvum* is a zoonotic species and the predominant in preweaned calves, especially those at age of 1-4 weeks <sup>[8]</sup>. The agent is responsible for about 85% of cryptosporidiosis in preweaned calves but only 1% of the disease in postweaned calves and 1-2 year old heifers <sup>[6,9,10]</sup>. Among the other bovine species, *C. bovis* and *C. ryanae* were detected mainly in weaned calves, and *C. andersoni* in yearlings and adult cattle <sup>[9,11,12]</sup>. While *C. bovis* and *C. ryanae* are considered non-zoonotic, *C. andersoni* has recently been reported in few research involving humans in England <sup>[13]</sup>.

The specific diagnosis of Cryptosporidium species is central to the control of the disease and to the understanding of the epidemiology. Lack of distinctive morphologic features of Cryptosporidium oocysts makes microscopical examination inconvenient in order to clearly differentiate species and genotypes [14]. Traditionally, C. parvum has been diagnosed by microscopy of fecal smears, with or without staining. However, two other species, C. bovis and C. ryanae, with similar oocyst morphology to C. parvum, can only be identified using DNA analysis. That is, microscopy cannot distinguish these three species <sup>[6]</sup>. Therefore, molecular analyses are required to detect and distinguish Cryptosporidium at species/genotype and subtype levels <sup>[3,14]</sup>. The most frequently used marker for Cryptosporidium species and genotype identification is the small subunit of ribosomal RNA (SSU-rRNA) gene<sup>[3]</sup>.

The disease in calves has been studied in many countries, with prevalence ranging from 2.4 to 100% <sup>[3,7]</sup>. In Turkey, cryptosporidiosis was first diagnosed in calves in 1984 <sup>[15]</sup>. Since then, other surveys have revealed the prevalence of 7.2-63.9% in calves <sup>[16,17]</sup>. Most of the studies carried out in Turkey were based on microscopy of stained oocysts in feces <sup>[15,16,18-21]</sup>. Enzyme-linked immunosorbent assay (ELISA) <sup>[17,22]</sup> and PCR technique <sup>[23-25]</sup> have recently become more common to attain the prevalence of cryptosporidiosis. However, few studies have coped with

genetic structure of *Cryptosporidium* species in Turkey <sup>[26-28]</sup>. This study was conducted to determine the prevalence of cryptosporidiosis and to characterize *Cryptosporidium* species based on PCR amplification and sequence analysis of SSU rRNA gene in younger than 1-month-old calves in Erzurum province, Turkey.

# **MATERIAL and METHODS**

### Sample Collection

A total of 307 fecal samples were collected from calves less than 1 month old in dairy farms (herd size ranging from 100 to 350 Brown Swiss, and Holstein cows in 3 professional dairy farms and from 40 to 85 Brown Swiss, crossbreed, and Anatolian Red cows in 2 traditional dairy farms) located in Erzurum province between April-2010 and October-2010. Samples were collected directly from the rectum with a gloved hand and transferred into a plastic cup. Fecal consistency was scored as firm, well formed, loose and diarrheic. Samples were kept at 4°C until laboratory analyses.

The study protocol was approved by the Animal Care and Use Committee at Ataturk University (4.4.2008-2008/8 decision number).

#### DNA Extraction and PCR Amplification

Oocysts were washed and concentrated from feces <sup>(29,30)</sup> prior to DNA isolation using a QIAamp DNA Stool kit (Qiagen, Maryland, USA). Before eluting, aliquots were added with 100 ml Buffer AE and stored at 20°C.

A nested PCR for the amplification of a fragment of SSU rRNA gene was performed using the protocols and primers as described by Xiao et al.[31] with the following modifications: At the first step of nested PCR, approximately 1.325 bp PCR product was amplified using primers 5'-TTCTAGAGCTAATACATGCG-3' and 5'-CCCATTTCCTTCGAAA CAGGA-3'. The PCR contained 1x PCR buffer, 6 mM MgCl<sub>2</sub>, 0.2 mM (each) dNTP, 200 nM (each) primer, 0.025 U of Tag DNA polymerase, and 1.5 µl of DNA template in a total 25 µl reaction mixture. A total of 35 cycles were carried out at 94°C for 45 s, 55°C for 45 s and 72°C for 1 min. There was also an initial hot start at 94°C for 3 min and a final extension at 72°C for 7 min. A secondary PCR was then performed to amplify 826-864 bp from 1 µl of the primary PCR mixture using primers 5'-GGAAGGGTTGTATTTATTAGATAAAG-3' and 5'-AAGGAGTAAGGAACAACCTCCA-3'. The PCR and cycling conditions were identical to the primary PCR. Amplification products were separated by electrophoresis on 1% (w/v) agarose gels, and visualized by ethidium bromide staining.

#### **DNA Sequence Analysis and Phylogenetic Analysis**

Successfully amplified samples were subjected to DNA sequence analysis for species determination. Sequencing

Table 1 Eactors affecting cryptosporidiosis in calves younger than one

was performed using the ABI PRISM® BigDye terminator cycle sequencing kit in ABI PRISM 310 genetic analyzer (Applied Biosystems, Foster City, CA). Sequence data were then subjected to BLASTN (RefSeq) searches of the Cryptosporidium genome database at the National Center for Biotechnology Information (http://www.ncbi nlm.nih. gov/). All sequence data were edited using BioEdit 7.0 (http: //www.mbio.ncsu.edu/BioEdit/bioedit.html) and FinchTV Version 1.4.0 (http://www.geospiza.com/finchtv) following naked eye checking. Multiple sequence alignments were made with the Clustal W method with BioEdit 7.0 software <sup>[32]</sup>. The neighbor-joining (NJ) method as implemented in the MEGA5.1 program <sup>[33]</sup> was used for the phylogenetic analysis based on SSU rRNA, utilizing Eimeria tenella sequence (HQ680474) as out-group. The branch reliability was assessed by the bootstrap method with 1000 replications.

#### **Data Analysis**

The PROC MEANS and FREQ were computed to obtain descriptive statistics <sup>[34]</sup>. Animals were categorized by breed (culture, crossbreed and local), age (1-7, 8-15 and 16-30 days) and fecal consistency (firm, well formed, loose and diarrheic) before establishing cross-tables to reveal association of risk factors with cryptosporidiosis using Chi-square. The associations were considered significant at P<0.05.

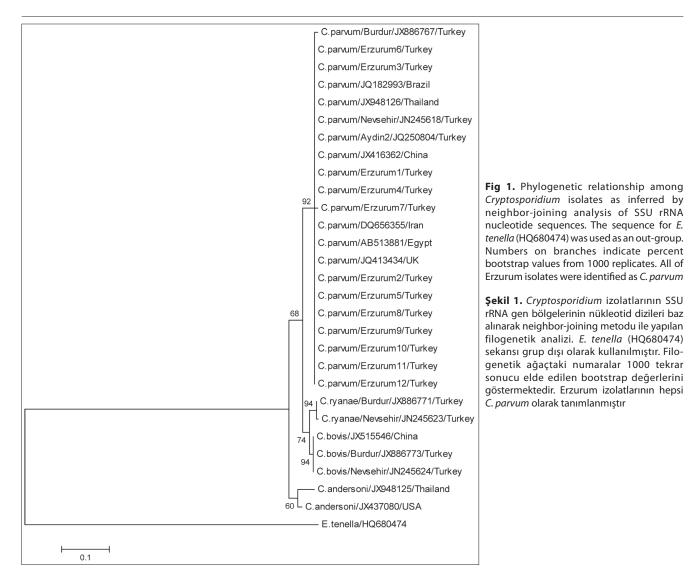
## RESULTS

The nested PCR showed a total of 12 (3.9%) positive amplications in 307 fecal samples (*Table 1*). The cryptosporidiosis prevalence among calves raised in modern farm was lower than those raised in farms with poor infrastructure (1.5 vs. 8.5%, P<0.003). The infection rate in culture breed was insignificantly higher (4.5%) than the other breeds. As the age advanced, the frequency of animals infected by *Cryptosporidium* increased quadratically, being the highest among calves aged 8-15 days (P<0.0001). Cryptosporidiosis was more common in females than males (6.8 vs. 1.3%, P<0.01). The cryptosporidiosis prevalence increased with the fecal water content (P<0.0001), 13.6% in calves with the diarrheic feces and 7.1% in calves with the loose feces (*Table 1*).

All PCR-positive isolates in Erzurum were confirmed to be *C. parvum (Fig. 1)*. The sequences were deposited into GenBank under accession numbers KC437395 to KC437406, respectively. *C. parvum* [GenBank: JN245618, JQ250804, JX886767, DQ656355, AB513881, JX948126, JX416362, JQ413434, JQ182993], *C. andersoni* [GenBank: JX948125, JX437080], *C. bovis* [GenBank: JX515546, JX886773, JN245624] and *C. ryanae* [GenBank: JX886771, JN245623] were reference species, whereas *E. tenella* (HQ680474) was an out-group reference in comparisons. *Fig.* 1 depicts phylogenetic relationship among *C. parvum* 

Variable	Infection Status	
	PCR – (n = 295, 96.09%)	PCR + (n = 12, 3.91%)
Enterprise	1	1
Traditional (n=106, 34.53%)	97 (91.5)	9 (8.5)
Professional (n=201, 65.47%)	198 (98.5)	3 (1.5)
	X <sup>2</sup> = 9.05, <i>P</i> <0.003	
Breed		
Culture (n=264, 85.99%)	252 (95.4)	12 (4.5)
Crossbreed (n=8, 2.61%)	8 (100)	0
Local (n=35, 11.40%)	35 (100)	0
	X <sup>2</sup> = 2.03, P<0.36	
Age (d)		
1-7 (n=30, 9.77%)	28 (93.3)	2 (6.7)
8-15 (n=71, 23.13%)	62 (87.3)	9 (12.7)
16-30 (n=206, 67.10%)	205 (99.5)	1 (0.5)
	X <sup>2</sup> = 21.56, P<0.0001	
Sex	1	1
Female (n=148, 48.21%)	138 (93.2)	10 (6.8)
Male (n=159, 51.79%)	157 (98.7)	2 (1.3)
	<i>X</i> <sup>2</sup> = 6.17, <i>P</i> <0.01	
Fecal consistency		
Firm (n=11, 3.58%)	11 (100)	0
Well formed (181, 58.96%)	181 (100)	0
Loose (n=56, 18.24%)	52 (92.9)	4 (7.1)
Diarrheic (n=59, 19.22%)	51 (86.4)	8 (13.6)
	X <sup>2</sup> = 24.00, P<0.0001	

in Erzurum isolates and the other *Cryptosporidium* isolates as inferred by the NJ analysis of the partial SSU rRNA gene sequences. *C. parvum* in Erzurum isolates were grouped into the same clade with respective reference *C. parvum* sequences. In the present experiment, the percent identities were 99.3-100% among *C. parvum* in Erzurum isolates, 98.7-100% with other *C. parvum* isolates and 87.8-94% with other *Cryptosporidium* species from GenBank.



## DISCUSSION

The prevalence of cryptosporidiosis in Turkey varies between 7.2-63.9% in calves <sup>[16,17]</sup>. To our knowledge, this study delivered the lowest prevalence rate (3.9%) among other reports from different locations of Turkey <sup>[17,19-22,24,26-28]</sup>. The difference could be due to a vast number of factors such as breed, age, management, environment, and season as well as diagnostic method <sup>[5,7,13]</sup>. The low prevalence could also be caused by spot fecal sampling instead of serial sampling, which may result in underestimation because of intermittent oocyst excretion <sup>[9,11]</sup>.

The majority of *C. parvum* infections appear to be limited to dairy calves under eight weeks of age <sup>[10,35]</sup>, being highest in calves up to 1-month-old <sup>[7,8,36]</sup>. In calves, the highest infection rates are reported in calves 7-14 days old <sup>[7,37]</sup>, 8-14 days old <sup>[4,38]</sup> and 8-21 days old <sup>[39]</sup>. In accordance with the literature, in the present study, the infection prevalence was highest in calves aged between 8-15 days (12.7%), followed by those aged 1-7 days (6.7%) and 16-30 days (0.5%).

As previously reported by Trotz-Williams et al.<sup>[40]</sup> in Ontario, Canada, by Aysul et al.<sup>[26]</sup> in Aydın, Turkey and by Coklin et al.<sup>[13]</sup> in Prince Edward Island, Canada, *C. parvum* was the only species identified in calves less than 1 month old. On the other hand, the absence of *C. bovis*, *C. andersoni* and *C. bovis* in our study could be a result of the age group ( $\leq$  1 months) because since *C. bovis* and *C. ryanae* are known to be more prevalent in weaned calves and *C. andersoni* in yearlings and adult cattle <sup>[6,9,11,12]</sup>.

Calf diarrhea has a multifactorial etiology, and *C. parvum* is frequently associated with the disease <sup>[7,38,39,41]</sup>. Besides, viruses and bacteria are other causative agents that can cause this symptom simultaneously or individually. Of 12 *C. parvum* positive fecal samples, 8 were from diarrheic calves and 4 from calves with loose feces (*Table 1*). In disagreement with some previous studies <sup>[35,39,41,42]</sup>, our results proved an association of fecal consistency with the infection. Studies reporting relationship between fecal consistency and cryptosporidiosis are available <sup>[7,12,36]</sup>. Because other possible agents were not searched in the present study, it requires caution to make inference that

calves with watery feces are prone to cryptosporidiosis. Another factor to contribute fecal dry matter is feeding scheme because looser feces can be consequence of milk feeding <sup>[39]</sup>. These suggest that extensive sample analysis is required to confirm the relationship between fecal consistency and cryptosporidiosis.

The molecular characterization of *Cryptosporidium* species in Turkey has been published in three reports, in which *C. parvum* <sup>[26-28]</sup>, *C. bovis* <sup>[27]</sup> and *C. ryanae* <sup>[27]</sup> were identified. In our study, homology search proved that all isolates in Erzurum were *C. parvum*. The partial SSU rRNA gene sequences had 100% similarity to reference sequences downloaded from the GenBank (DQ656355, AB513881, JX948126, JX416362, JQ413434, JQ182993 and JN245618). The NJ phylogenetic analysis based on the SSU rRNA (*Fig. 1*) showed that all sequences of *C. parvum* in Erzurum isolates clustered in the intestinal clade with reference *C. parvum* sequences (bootstrap value 92).

In conclusion, the current study elucidated the prevalence of cryptosporidiosis and the molecular characterization of *Cryptosporidium* species found in calves in Erzurum, Turkey. The prevalence of *Cryptosporidium* infection in dairy calves determined by nested PCR was at 3.9%. *C. parvum* was the only causative *Cryptosporidium* species in calves younger than 1 month in Erzurum province as ascertained by sequencing the amplified SSU rRNA regions.

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#### REFERENCES

1. Fredes F, Díaz A, Raffo E, Munoz P: Cryptosporidium spp. oocysts detected using acid-fast stain in feces of gentoo penguins (*Pygoscelis papua*) in Antarctica. Antarct Sci, 20, 495-496, 2008.

**2. Fayer R:** Taxonomy and species delimitation in *Cryptosporidium. Exp Parasitol*, 124, 90-97, 2010.

**3. Xiao L:** Molecular epidemiology of cryptosporidiosis: An update. *Exp Parasitol*, 124, 80-89, 2010.

**4. Imre K, Lobo LM, Matosb O, Popescu C, Genchid C, Darabus G:** Molecular characterisation of *Cryptosporidium* isolates from pre-weaned calves in Romania: Is there an actual risk of zoonotic infections? *Vet Parasitol*, 181, 321-324, 2011.

**5. Venu R, Latha BR, Basith SA, Raj GD, Sreekumar C, Raman M:** Molecular prevalence of *Cryptosporidium* spp. in dairy calves in Southern states of India. *Vet Parasitol*, 188, 19-24, 2012.

**6. Silverlås C, Näslund K, Björkman C, Mattsson G:** Molecular characterization of *Cryptosporidium* isolates from Swedish dairy cattle in relation to age, diarrhoea and region. *Vet Parasitol*, 169, 289-295, 2010.

7. Díaz-Lee A, Mercado R, Onuoha EO, Ozaki LS, Munoz P, Munoz V, Martínez FJ, Fredes F: *Cryptosporidium parvum* in diarrheic calves detected by microscopy and identified by immunochromatographic and molecular methods. *Vet Parasitol*, 176, 139-144, 2011.

**8. Starkey SR, Wade SE, Schaaf S, Mohammed HO:** Incidence of *Cryptosporidium parvum* in the dairy cattle population in a New York City watershed. *Vet Parasitol*, 131, 197-205, 2005.

9. Santín M, Trout JM, Fayer R: A longitudinal study of cryptosporidiosis in

dairy cattle from birth to two years of age. *Vet Parasitol*, 155, 15-23, 2008. **10. Brook E, Hart CA, French N, Christley R:** Molecular epidemiology of *Cryptosporidium* subtypes in cattle in England. *Vet J*, 179, 378-382, 2009.

**11. Fayer R, Santín M, Trout JM, Greiner E:** Prevalence of species and genotypes of *Cryptosporidium* found in 1-2-year-old dairy cattle in the eastern United States. *Vet Parasitol*, 135, 105-112, 2006.

**12. Fayer R, Santín M, Trout JM:** Prevalence of *Cryptosporidium* species and genotypes in mature dairy cattle on farms in eastern United States compared with younger cattle from the same locations. *Vet Parasitol*, 145, 260-266, 2007.

**13.** Coklin T, Uehlinger FD, Farber JM, Barkema HW, O'Handley RM, Dixon BR: Prevalence and molecular characterization of *Cryptosporidium* spp. in dairy calves from 11 farms in Prince Edward Island, Canada. *Vet Parasitol*, 160, 323-326, 2009.

**14.** Jex AR, Smith HV, Monis PT, Campbell BE, Gasser RB: *Cryptosporidium*-Biotechnological advances in the detection, diagnosis and analysis of genetic variation. *Biotechnol Adv*, 26, 304-317, 2008.

**15. Burgu A:** Türkiye'de buzağılarda *Cryptosporidium*'ların bulunuşu ile ilgili ilk çalışmalar. *Ankara Üniv Vet Fak Derg*, 31 (3): 573-585, 1984.

**16. Özer E, Erdoğmuş SZ, Köroğlu E:** Elazığ yöresinde buzağı ve kuzularda bulunan *Cryptosporidium*'un yayılışı üzerinde araştırmalar. *Doğa Turk J Vet Anim Sci*, 14, 439-445, 1990.

**17. Sevinç F, Irmak K, Sevinç M:** The prevalence of *Cryptosporidium parvum* infection in the diarrhoeic and non-diarrhoeic calves. *Revue Med Vet*, 154 (5): 357-361, 2003.

**18. Özlem MB, Eren H, Kaya O:** Aydın yöresi buzağılarında *Cryptosporidium'* ların varlığının araştırılması. *Bornova Vet Kontr Araşt Enst Md Derg*, 22 (36): 15-22, 1997.

**19. Değerli S, Çeliksöz A, Kalkan K, Özçelik S:** Prevalence of *Cryptosporidium* spp. and *Giardia* spp. in cows and calves in Sivas. *Turk J Vet Anim Sci*, 29, 995-999, 2005.

**20. Göz Y, Gül A, Aydın A:** Hakkari yöresinde sığırlarda *Cryptosporidium* sp.'nin yaygınlığı. *YYÜ Vet Fak Derg*, 18, 37-40, 2007.

21. Aştı C, Özbakış G, Azrug AF, Orkun Ö, Nalbantoğlu S, Çakmak A, Burgu A: Farklı illere ait buzağı dışkı bakısı sonuçları. *Kafkas Univ Vet Fak Derg*, 18 (Suppl-A): A209-A214, 2012.

**22. Çiçek M, Körkoca H, Gül A:** Van belediyesi mezbahasında çalışan işçilerde ve kesimi yapılan hayvanlarda *Cryptosporidium* sp.'nin araştırılması. *T Parazitol Derg*, 32 (1): 8-11, 2008.

**23. Arslan MÖ, Erdoğan HM, Tanrıverdi S:** Neonatal buzağılarda Cryptosporidiosis'in epidemiyolojisi. *13. Ulusal Parazitoloji Kongresi*, *Program ve Özet Kitabı*, SB6-01, s. 186, 8-12 Eylül, Konya-TÜRKİYE, 2003.

**24.** Sungur T, Kar S, Güven E, Aktaş M, Karaer Z, Vatansever Z: *Cryptosporidium* spp'nin dışkıdan nested-PCR ve carbol fuchsin boyama yöntemi ile teşhis edilmesi. *T Parazitol Derg*, 32 (4): 305-308, 2008.

**25.** Sakarya Y, Kar S, Tanyüksel M, Karaer Z, Babür C, Vatansever Z: Detection of *Cryptosporidium* spp. in humans and calves through nested PCR and carbol fuchsin staining methods in Ankara, Turkey. *Kafkas Univ Vet Fak Derg*, 16 (6): 977-980, 2010.

**26. Aysul N, Ulutaş B, Ünlü H, Hoşgör M, Atasoy A, Karagenç T:** Aydın ilinde ishalli buzağılarda bulunan *Cryptosporidium* türlerinin moleküler karakterizasyonu. *16. Ulusal Parazitoloji Kongresi, Program ve Özet Kitabı,* S-15, s. 208, 1-7 Kasım, Adana-TÜRKİYE, 2009.

27. Şimşek AT, İnci A, Yıldırım A, Çiloğlu A, Bişkin Z, Düzlü Ö: Nevşehir yöresinde ishalli buzağılarda *Cryptosporidium* türlerinin moleküler prevalansı ve karakterizasyonu. *17. Ulusal Parazitoloji Kongresi, Program ve Özet Kitabı*, SB03-06, s.158, 5-10 Eylül, Kars-TÜRKİYE, 2011.

**28. Arslan MÖ, Ekinci Aİ:** Kars yöresinde sığırlarda *Cryptosporidium parvum* subtiplerinin belirlenmesi. *Kafkas Univ Vet Fak Derg*, 18 (Suppl-A): A221-A226, 2012.

**29. Fayer R, Morgan U, Upton SJ:** Epidemiology of *Cryptosporidium*: transmission, detection and identification. *Int J Parasitol*, 30, 1305-1322, 2000.

30. Santín M, Trout JM, Xiao L, Zhou L, Greiner E, Fayer R: Prevalence and age-related variation of *Cryptosporidium* species and genotypes in dairy calves. Vet Parasitol, 122, 103-117, 2004.

**31. Xiao L, Singh A, Limor J, Graczyk TK, Gradus S, Lal A:** Molecular characterization of *Cryptosporidium* oocysts in samples of raw surface water and wastewater. *Appl Environ Microb*, 67, 1097-1101, 2001.

**32. Thompson JD, Gibson TJ, Plewniak F, Jeanmougin F, Higgins DG:** The Clustal X windows interface: Flexible strategies for multiple sequences alignment aided by quality analysis tools. *Nucleic Acids Res*, 25, 4876-4882, 1997.

**33. Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S:** MEGA5: Molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance and maximum parsimony methods. *Mol Biol Evol*, 28, 2731-2739, 2011.

**34. SAS:** Statistical analysis system, User's Guide, Version 9. SAS Institute Inc, Cary, NC, 2002.

**35. Silverlås C, Bosaeus-Reineck H, Näslund K, Björkman C:** 2012. Is there a need for improved *Cryptosporidium* diagnostics in Swedish calves? *Int J Parasitol*, 43 (2): 155-161, 2013.

**36.** Maurya PS, Rakesh RL, Pradeep B, Kumar S, Kundu K, Garg R, Ram H, Kumar A, Banerjee PS: Prevalence and risk factors associated with *Cryptosporidium* spp. infection in young domestic livestock in India. *Trop Anim Health Prod*, 45 (4): 941-946, 2013.

**37. Tanriverdi S, Markovics A, Arslan MO, Itik A, Shklap V, Widmer G:** Emergence of distinct genotypes of *Cryptosporidium parvum* in structured host populations. *Appl Environ Microbiol*, 72, 2507-2513, 2006.

**38. de la Fuente R, Luzon M, Ruiz-Santa-Quiteria JA, Garcia A, Cid D, Orden JA, Garcia S, Sanz R, Gomez-Bautista M:** *Cryptosporidium* and concurrent infections with other major enterophatogens in 1 to 30-day-old diarrheic dairy calves in central Spain. *Vet Parasitol*, 80, 179-185, 1999.

**39.** Brook E, Hart CA, French N, Christley R: Prevalence and risk factors for *Cryptosporidium* spp. infection in young calves. *Vet Parasitol*, 152, 46-52, 2008.

40. Trotz-Williams LA, Martin DS, Gatei W, Cama V, Peregrine AS, Martin SW, Nydam DV, Jamieson F, Xiao L: Genotype and subtype analyses of *Cryptosporidium* isolates from dairy calves and humans in Ontario. *Parasitol Res*, 99, 346-352, 2006.

**41. Silverlås C, de Verdier K, Emanuelson U, Mattsson JG, Björkman C:** *Cryptosporidium* infection in herds with and without calf diarrheal problems. *Parasitol Res*, 107, 1435-1444, 2010.

**42. Maikai BV, Umoh JU, Kwaga JKP, Lawal IA, Maikai VA, Cama V, Xiao** L: Molecular characterization of *Cryptosporidium* spp. in native breeds of cattle in Kaduna State, Nigeria. *Vet Parasitol*, 178, 241-245, 2011.